

Evaluate the Ability of Environment isolates of *Bacillus licheniformis* to Synthesized Gold Nanoparticles

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Abstract

The current study aimed to investigate the ability of *Bacillus licheniformis* isolated from the environment to synthesize gold nanoparticles (AuNPs). The study included isolating and identifying of *B. licheniformis* from the soil and investigating its ability to synthesis AuNPs, then identifying AuNPs using XRD, UV -vis., FeSEM, EDS, FTIR, and AFM. The results showed the ability of *B. licheniformis* isolated from soil to synthesis AuNPs by changing the color of the solution from yellow to ruby pink and the optical absorption using ultraviolet spectrometry showed the appearance of a beak of AuNPs at a wavelength of 530 nm. FeSEM electron microscopy showed that synthesized AuNPs were spherical in shape and homogeneous with a diameter ranging from 20.47 - 76.50 nm. Also, the results of XRD, FTIR, EDS, and AFM revealed the nanoscale properties of AuNPs synthesized from *B. licheniformis*.

Key Words: AuNPs, *B. licheniformis*, XRD, FTIR, EDS, AFM.

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INTRODUCTION

Gold nanoparticles (AuNPs) is one of the most noble metals had been studied due to their unique properties and functionalities of surface that can be used in many aspects including biosensors, microbiology, pharmacy, biotechnology, catalysis, ect. (Rahi et al., 2015). Surfaces of AuNPs can be modified readily with ligands that contain the functional groups such as phosphines, thiols, and amines which exhibit affinity for gold surfaces (Mahato et al., 2019). Due to the wide range of AuNPs and biocidal properties, Au-based compound has been utilized as nontoxic inorganic agents, its compounds are non-toxic to cells

of animal but toxic to microorganisms such as fungi and bacteria (Sayadi et al., 2021). AuNPs come in a variety of sizes and shapes such as spherical, decahedral, icosahedral multiple twined, sub-octahedral, nanoprisms, octahedral, nanotriangles, irregular shape, tetrahedral, hexagonal platelets and nanorods (A. K. Khan et al., 2014). Physical, chemical, and biological approaches were used to synthesize metal nanoparticles. Chemical and physical processes give higher yields, but they require specialized and expensive equipment, high energy consumption, difficult circumstances, in addition, these nanoparticles (NPs) cannot be used in the medicine because of health-related issue (Alzubaidi et al., 2023; Huq et al., 2022). Some Nanoparticles had been synthesized by such as silver nanoparticles which produced by *B. licheniformis* showed antimicrobial activity against human pathogens including gram negative and gram positive bacteria, fungi and viruses (Elbeshehy et al., 2015). Silver and Gold Nanoparticles were produced by using *B. licheniformis* (R. Singh et al., 2014; Sriram et al., 2012) and cadmium nanoparticles (Bakhshi & Hosseini, 2016). selenium nanoparticles derived biogenic that produced by *B. licheniformis* had been reported as new prevention strategy reducing prostate cancer (Sonkusre, 2020). *B. licheniformis* have ability to produce several including proteases, α -amylase, β -mannanase, pentosanase, cycloglucosyl transferase, penicillinase, and several pectinolytic enzyme (Rey et al., 2004).

METHODS

Isolation and Identification of *B. licheniformis*

Fifteen soil samples were collected from different sites including: garden, barren and

river banks during November 2021. The soil samples were collected from 10 cm depth in a sterile screw caps and suspended in 9 ml of distilled water, mixed well, diluted and the second tube (10-2) was placed in water bath at 80 °C for 10 min in order to kill all vegetative cells remaining only spores forming bacteria (Organji et al., 2015). One ml of the bacterial suspension was transferred to tubes containing 9 ml of BHI broth and incubated at 37°C for 24-48 hours. After incubation a swab from each positive tubes (tubes with turbidity) were transferred to nutrient agar and blood agar plates for the isolation and primary identification. In addition to morphological character of bacterial colonies and microscopic examination, a confirmed identification of *B. licheniformis* was carried out using vitek 2 compact system.

Synthesized of AuNPs by *B. licheniformis*

The approach described by Singh & Kundu, 2014, has been followed to assess extracellular production of AuNPs. Purified bacterial isolates were grown in 100 mL of nutrient broth for 24 hours at 37° C for biomass generation. The culture was centrifuged for 30 minutes at 8000 rpm/min to separate the bacterial cells. In a new sterile conical flask, the supernatant was collected. One mM of HAuCl₄.3H₂O (chlorouric acid trihydrates) has been added to 5 ml of the all isolates supernatant and incubated in a shaker at 37°C for 24 hours, the ability of bacterial supernatant to create AuNPs was assessed.

Characterization of AuNPs

Only isolates that showed high ability to synthesis AuNPs were characterized by UV visible, SEM, XRD, EDS, and FTIR. All these technique were carried out in the Biological Center of Tahrán University / Iran, except UV visible characterized which carried out in Alameen Center For Advanced Research and Biotechnology (ACARB) in Najaf / Iraq.

RESULTS AND DISCUSSION

The results of primary isolation of *B. licheniformis* was shown in figure (1). Morphologically, on blood agar base its appear as opaque white colonies with a coarse and matte surface with irregular edge with no hemolytic activity while it

appearance in nutrient agar was large opaque, adherent and irregular edges this was proven by researchers (Ghani et al., 2013). Microscopically, it is appear as a rod-shaped Gram-positive cells, arranged in diploid form or long chains. Vitek 2 Compact System confirmed the identification of *B. licheniformis* as mention in table 1.

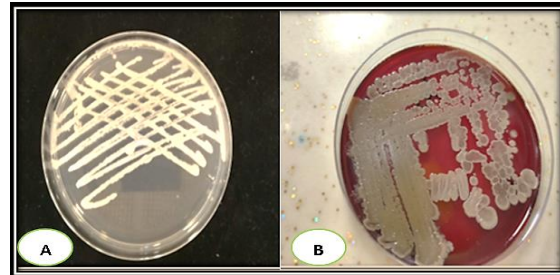


Fig.1. *Bacillus licheniformis* isolate on A: nutrient agar, B: blood agar base .

Table.1. The result of the vitek 2 compact system for identification of *Bacillus licheniformis*.

BioMérieux Customer:		Laboratory Report		Printed by: LabAdmin Patient ID:							
System #:		Card:	BCL	Lot Number:	2391888103						
Patient Name:		Status:	Final	Analysis Timer:	13:55 hours						
Isolate: Ss-1 (Qualified)		Organism Origin:	VITEK 2	Expires:	Jun 27, 2023 12:00 CST						
Card Type: BCL Bar Code: 2391888103095398		Selected Organism:	<i>Bacillus licheniformis</i>	Complete:	Jun 15, 2020 21:22 CDT						
Setup Technology: Laboratory Administrator(LabAdmin)		Biomass:	85% Probability								
Biomnumber: 0332371514466221		Confidence:	Acceptable identification								
Organism Quantity:		Analysis Messages:									
		Contradicting Typical Biopatterns:									
		<i>Bacillus licheniformis</i>	APP(1),TTZ(94),Gly(A)S3,AGLU(99),PLE(97),								
Biochemical Details											
1	BXYL	3	LysA	4	AspA	5	LeuA	7	PheA	8	ProA
9	BGAL	10	ProA	11	AGAL	12	AlbA	13	TyrA	14	BNAG
15	APPA	18	COXA	19	BGAL	21	GlyG	22	IND	24	MSG
25	ELLM	26	MDX	27	AMAN	29	MTE	30	GlyA	31	dMAN
32	dMNE	34	dMLZ	36	NAG	37	PLE	39	IRHA	41	BGLU
43	BMAN	44	PHC	45	PVATE	46	AGLU	47	dTAG	48	dTRE
50	INU	53	dGLU	54	dRH	56	PSCNa	58	NaCl	59	KAN
60	OLD	61	ESC	62	TTZ	63	POLYB	65	NaCl	6.5%	

Biosynthesis of AuNPs

A large amount of AuNPs have been synthesized by *B. licheniformis*, the results showed the efficiency of isolate to synthesis AuNPs by changing the color of bacterial supernatant from yellow to pink-ruby (Figure 2).



Fig.2. Biosynthesis of Gold nanoparticles by *Bacillus licheniformis* showed the color variation from yellow to pink-ruby.

CHARACTERIZATION OF SYNTHESIZED AUNPS

UV-visible spectroscopic examination

The results of UV-visible spectroscopic showed that the absorption peak of AuNPs synthesized by *B. licheniformis* was at a wavelength of approximately 520 nm. Change the color of bacterial suspension from yellow to red and measurement of the absorbance bands using UV-visible spectroscopy can confirm the biogenesis of AuNPs by *B. licheniformis* (Figure 3). The UV-visible spectrophotometric method is a true and tried method for identifying nanoparticles. The color of reaction mixture changed after 24 hours of incubation which indicating the synthesis of nanoparticles in the mixture. The AuNPs synthesized using bacteria produced SPR values in the range of 400–700 nm (Menon et al., 2017; Mussa, 2019) or (500-600) (Diego et al., 2018). As time progresses, there is a possibility of aggregation of NPs which results in a shift of longer wavelength region (Soliman et al., 2022). AuNPs synthesized by plant extract showed an absorption band at 225 nm (Rotimi et al., 2019). While that synthesized by *Bacillus subtilis* bacteria was 545nm (Srinath et al., 2018).

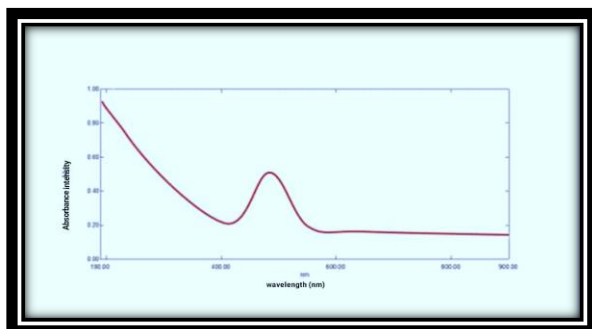


Fig.3. UV-VIS spectrum of gold nanoparticles synthesis by *Bacillus licheniformis*.

Field Emission-Scanning Electron Microscopy (FE-SEM) Analysis

The results of FE-SEM of AuNPs synthesized by *B. licheniformis* showed a homogenous in diameter and spherical shapes nanoparticles were the diameter of AuNPs ranges between 20.47-76.50 with average 33.6 nm as showed in figure (4). Many of studies showed the spherical shape

of AuNPs (Yuan et al., 2023; Zhao et al., 2022; Depciuch et al., 2020). It has been confirmed that the reduction of biomolecules by mineral salts results in spherical nanoparticles (Sun et al., 2019)The variation in the shape and size of synthesized AuNPs may due to differences preparation method which lead to different shape and size of AuNPs (Gu et al., 2021).

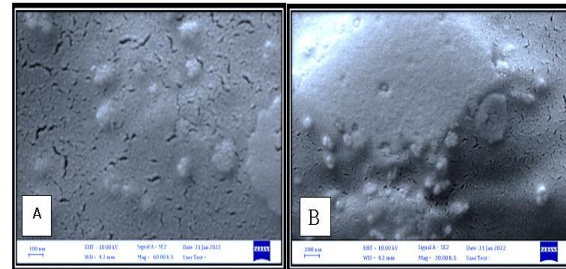


Fig.4. FE-SEM examination of gold nanoparticles synthesized by *Bacillus licheniformis* at: A: 100 nm, B: 200 nm.

X-Ray Diffraction (XRD) Analysis of AuNPs

As mention in figure 5, the results showed that the extracellular AuNPs synthesized by *B. licheniformis* are greatly crystalline in nature where four distinct peaks at 2θ values of 64.5, 44.1, 37.8 and 77.5 corresponding to 111, 200, 220, and 311 planes of AuNPs synthesis by *B. licheniformis*. XRD spectra of pure crystalline Au structures was published by the Joint Committee on Powder Diffraction Standards (file nos. 04-0784). The size of these AuNPs were 34.5 nm according XRD analysis. These results are compatible reported findings of characterization of AuNPs (Fereig et al., 2022; Xie, et al., 2019; Rajeshkumar, 2016).

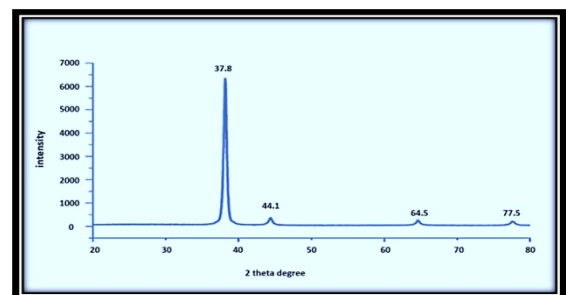


Fig.5. XRD of gold nanoparticles synthesis by *Bacillus licheniformis*.

Fourier Transform Infrared (FTIR) Analysis

The results of FTIR chart (Figure 6) showed the appearance of different peaks at range between 500-4000 cm^{-1} . The occurrence of

different peaks of absorption referred to the presence of other chemical groups such as O-H stretching group, C-H stretching group of alkane and N-H group. As mention in table (2) the peak of absorption of carboxylic group occur at 3911.5-3375.90 cm⁻¹ and 3448.80 cm⁻¹ which referred to the present of carboxylic group in carbohydrate, proteins amino acid residue that accumulated during the process of reduction of gold ions for synthesis of AuNPs (Diego et al., 2018). The presence of different functional groups in the AuNPs synthesized by bacteria indicate the role of these proteins in the synthesis and stabilization (preventing agglomeration) of the nanoparticles where the original proteins and the mechanisms of synthesis are not clear (San Diego et al., 2021). (Rabeea et al., 2020) suggest that the FTIR explain the attachments of polysaccharides, phenol and amino acids as reducer agents on AuNPs. The materials used in the manufacture of nanoparticles often contain a wide range of biologically active substances, which makes it difficult to determine the exact component responsible for the synthesis (F. Khan et al., 2021).

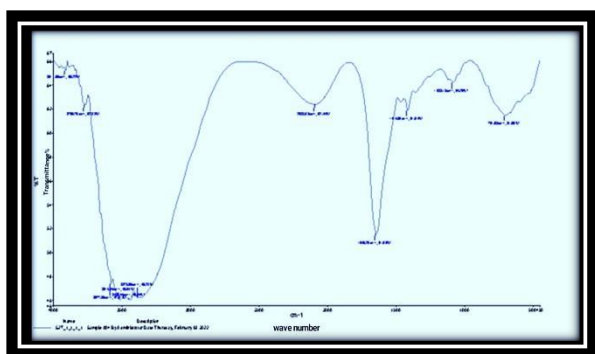


Fig.6. Fourier transform infrared spectroscopy (FTIR) of Nano biosynthesized Gold Nanoparticles by *Bacillus licheniformis*.

Other peaks have shown in this study at 1645.26- 1083.52 cm⁻¹ referred to carbonyl stretch and N-H bands vibration. N-H band vibration represent indicators of amid I and amid II groups of polypeptides as indicate by Faghihzadeh et al., 2016

Energy dispersive spectroscopy (EDS)

EDS analysis was used to define the crystal element of the presence of AuNPs by observed the optical absorption peaks. the element mapping visibly show the eminent amount of gold elements in the samples (Table 3). The results of EDS showed that the percentage of nanoparticles synthesized

by *B. licheniformis* were higher (40.71%) than other elements present in solution where the lower percentage of oxygen (9.3%) was observed. The percentages of other elements were as follow: C = 19.2%, Cl = 16.1% and Na = 14.7% (Figure 7 and Table 2). high amount of AuNPs with low and moderate amount of other elements that accompany the presence of the gold element may be produced from the components of a chemical generator used to adjust the value of the broth (Gupta & Padmanabhan, 2018) or may originated from bacterial biomolecules bound to the surface of AuNPs (Murugan et al., 2014)

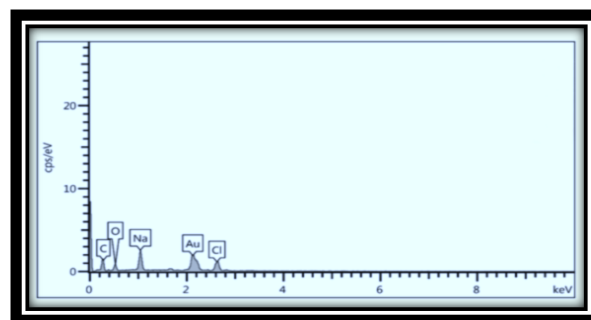


Fig.7. EDS analysis of gold nanoparticles synthesis by *Bacillus licheniformis*.

Table.2. The Percentage of Elements of gold nanoparticles synthesis by *Bacillus licheniformis*.

Elements	%
Au	40.7
C	19.2
Cl	16.1
Na	14.7
O	9.3
Total	100

ATOMIC FORCE MICROSCOPE (AFM)

The results of AFM analysis of AuNPs that synthesized by *B. licheniformis* showed (Figure 8) that AuNPs synthesized by *B. licheniformis* have physical size (6.397 × 6.397 μm) with average roughness about 4.294 nm. The rough surface depicts the synthesis of AuNPs. The hump depicts the big sized particles or the agglomerated AuNPs which fuse to give various exotic shapes in the later phase of reaction whereas the small individual peaks exposed in the images are of single spherical AuNPs. SEM measurements supported the AFM results about the structural features and

surface morphology of AuNPs synthesized well-dispersed spherical shaped. homogeneous population nanoparticles had arranged with regular surface shape with the peaks towards the top. The results showed homogenous in diameter and spherical shapes nanoparticles. The different preparation methods lead to different nanoparticle sizes and size distributions (Gu et al., 2021) These results are almost identical to (R. Singh et al., 2014) who got a size of AuNPs with average 33.6 nm and match with the results of the researchers (Gupta & Padmanabhan, 2018; Khalid et al., 2020). In the other study the AuNPs synthesized by *B. licheniformis* were nanocubes with range of size 10-100 (Kalishwaralal et al., 2009).

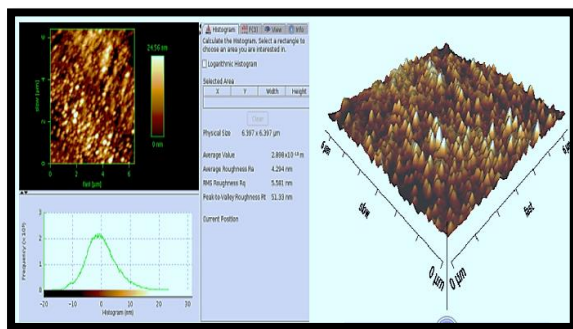


Fig.8. Atomic Force Microscopy (AFM) images of synthesized gold nanoparticles by *Bacillus licheniformis*.

CONCLUSION

The results showed the ability of *B. licheniformis* isolated from soil to synthesis AuNPs by changing the color of the solution from yellow to ruby pink and the optical absorption using ultraviolet spectrometry showed the appearance of a beak of AuNPs at a wavelength of 530 nm.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

FUNDING SOURCE

None

REFERENCES

1. Alzubaidi, A. K., Al-Kaabi, W. J., Ali, A. Al, Albukhaty, S., Al-Karagoly, H., Sulaiman, G. M., Asiri, M., & Khane, Y. (2023). Green synthesis and characterization of silver nanoparticles using flaxseed extract and evaluation of their antibacterial and antioxidant activities. *Applied Sciences*, 13(4), 2182.
2. Bakhshi, M., & Hosseini, M. R. (2016). Synthesis of CdS nanoparticles from cadmium sulfate solutions

3. Depciuch, J., Stec, M., Kandler, M., Baran, J., & Parlinska-Wojtan, M. (2020). From spherical to bone-shaped gold nanoparticles—Time factor in the formation of Au NPs, their optical and photothermal properties. *Photodiagnosis and Photodynamic Therapy*, 30, 101670.
4. Diego, K. D. G. S., Alindayu, J. I. A., & Baculi, R. Q. (2018). Biosynthesis of gold nanoparticles by bacteria from hyperalkaline spring and evaluation of their inhibitory activity against pyocyanin production. *Journal of Microbiology, Biotechnology and Food Sciences*, 8(2), 781–787. <https://doi.org/10.15414/jmbfs.2018.8.2.781-787>
5. Dziedzic, D. S. M., Mogharbel, B. F., Irioda, A. C., Stricker, P. E. F., Woiski, T. D., Machado, T. N., Bezerra Jr, A. G., & Athayde Teixeira de Carvalho, K. (2023). Laser Ablated Albumin Functionalized Spherical Gold Nanoparticles Indicated for Stem Cell Tracking. *Materials*, 16(3), 1034.
6. Elbeshehy, E. K. F., Elazzazy, A. M., & Aggelis, G. (2015). Silver nanoparticles synthesis mediated by new isolates of *Bacillus* spp., nanoparticle characterization and their activity against Bean Yellow Mosaic Virus and human pathogens. *Frontiers in Microbiology*, 6, 453.
7. Faghizadeh, F., Anaya, N. M., Schiffman, L. A., & Oyanedel-Craver, V. (2016). Fourier transform infrared spectroscopy to assess molecular-level changes in microorganisms exposed to nanoparticles. *Nanotechnology for Environmental Engineering*, 1(1). <https://doi.org/10.1007/s41204-016-0001-8>
8. Fereig, S., El-Zaafarany, G. M., Arafa, M., & Abdel-Mottaleb, M. M. A. (2022). Boosting the anti-inflammatory effect of self-assembled hybrid lecithin–chitosan nanoparticles via hybridization with gold nanoparticles for the treatment of psoriasis: elemental mapping and in vivo modeling. *Drug Delivery*, 29(1), 1726–1742. <https://doi.org/10.1080/10717544.2022.2081383>
9. Ghani, M., Ansari, A., Aman, A., Zohra, R. R., Siddiqui, N. N., & Qader, S. A. U. (2013). Isolation and characterization of different strains of *Bacillus licheniformis* for the production of commercially significant enzymes. *Pakistan Journal of Pharmaceutical Sciences*, 26(4), 691–697.
10. Gu, X., Xu, Z., Gu, L., Xu, H., Han, F., Chen, B., & Pan, X. (2021a). Preparation and antibacterial properties of gold nanoparticles: a review. *Environmental Chemistry Letters*, 19(1), 167–187. <https://doi.org/10.1007/s10311-020-01071-0>
11. Gu, X., Xu, Z., Gu, L., Xu, H., Han, F., Chen, B., & Pan, X. (2021b). Preparation and antibacterial properties of gold nanoparticles: a review. In *Environmental Chemistry Letters* (Vol. 19, Issue 1, pp. 167–187). Springer Science and Business Media Deutschland GmbH. <https://doi.org/10.1007/s10311-020-01071-0>
12. Gupta, R., & Padmanabhan, P. (2018). Biogenic synthesis and characterization of gold nanoparticles by a novel marine bacteria *Marinobacter algicola*: Progression from nanospheres to various geometrical shapes. *Journal of Microbiology, Biotechnology and Food Sciences*, 8(1), 732–737. <https://doi.org/10.15414/jmbfs.2018.8.1.732-737>
13. Huq, M. A., Ashrafudoulla, M., Rahman, M. M., Balusamy, S. R., & Akter, S. (2022). Green Synthesis and Potential Antibacterial Applications of Bioactive Silver Nanoparticles: A Review. *Polymers*, 14(4), 1–22. <https://doi.org/10.3390/polym14040742>
14. Kalishwaralal, K., Deepak, V., Ram Kumar Pandian, S., & Gurunathan, S. (2009). Biological synthesis of gold nanocubes from *Bacillus licheniformis*. *Bioresource Technology*, 100(21), 5356–5358. <https://doi.org/10.1016/j.biortech.2009.05.051>

15. Khalid, S., Parveen, S., Shah, M. R., Rahim, S., Ahmed, S., & Imran Malik, M. (2020). Calixarene coated gold nanoparticles as a novel therapeutic agent. *Arabian Journal of Chemistry*, 13(2), 3988–3996. <https://doi.org/10.1016/j.arabcj.2019.04.007>
16. Khan, A. K., Rashid, R., Murtaza, G., & Zahra, A. (2014). Gold nanoparticles: Synthesis and applications in drug delivery. *Tropical Journal of Pharmaceutical Research*, 13(7), 1169–1177. <https://doi.org/10.4314/tjpr.v13i7.23>
17. Khan, F., Kang, M. G., Jo, D. M., Chandika, P., Jung, W. K., Kang, H. W., & Kim, Y. M. (2021). Phloroglucinol-gold and-zinc oxide nanoparticles: Antibiofilm and antivirulence activities towards *Pseudomonas aeruginosa* pao1. *Marine Drugs*, 19(11), 1–24. <https://doi.org/10.3390/md19110601>
18. Hussain, M. A. ., Dawod , K. M. ., & Khether, A. A. . (2021). Gene Action, Heterosis and Combining Ability in Maize Hybrids B- Using Line x Tester Analysis. *Kufa Journal for Agricultural Sciences*, 13(2), 30–40. Retrieved from <https://journal.uokufa.edu.iq/index.php/kjas/article/view/3653>
19. Mahato, K., Nagpal, S., Shah, M. A., Srivastava, A., Maurya, P. K., Roy, S., Jaiswal, A., Singh, R., & Chandra, P. (2019). Gold nanoparticle surface engineering strategies and their applications in biomedicine and diagnostics. *3 Biotech*, 9(2), 1–19. <https://doi.org/10.1007/s13205-019-1577-z>
20. Menon, S., S., R., & S., V. K. (2017). A review on biogenic synthesis of gold nanoparticles, characterization, and its applications. *Resource-Efficient Technologies*, 3(4), 516–527. <https://doi.org/10.1016/j.refit.2017.08.002>
21. Murugan, M., Anthony, K. J. P., Jeyaraj, M., Rathinam, N. K., & Gurunathan, S. (2014). Biofabrication of gold nanoparticles and its biocompatibility in human breast adenocarcinoma cells (MCF-7). *Journal of Industrial and Engineering Chemistry*, 20(4), 1713–1719. <https://doi.org/10.1016/j.jiec.2013.08.021>
22. Mussa, L. A. M. (2019). Biosynthesis of Gold Nanoparticles by some Members of Enterobacteriaceae as Antibacterial Agent. PhD thesis, Faculty of Science, University of Babylon. Iraq.
23. Mohammed, D. ., Enouz, A. J. ., & Areaaer, A. H. J. . (2021). Effect of Adding Different Levels of Antioxidant and Imported Ginseng (Panax) Roots to The Diet in The Microbial . *Kufa Journal for Agricultural Sciences*, 13(2), 1–5. <https://doi.org/10.36077/kjas/2021/130201>
24. Organji, S. R., Abulreesh, H. H., Elbanna, K., Osman, G. E. H., & Khider, M. (2015). Occurrence and characterization of toxigenic *Bacillus cereus* in food and infant feces. *Asian Pacific Journal of Tropical Biomedicine*, 5(7), 515–520. <https://doi.org/10.1016/j.apjtb.2015.04.004>
25. Jeza, G. T., & Bekele, A. (2023). Seasonal distribution model of African elephants (*Loxodonta africana*) under a changing environment and land use in Omo National Park, Ethiopia. *Journal of Wildlife and Biodiversity*, 7(3), 96–117. <https://doi.org/10.5281/zenodo.7783039>
26. Rabeea, M. A., Owaid, M. N., Aziz, A. A., Jameel, M. S., & Dheyab, M. A. (2020). Mycosynthesis of gold nanoparticles using the extract of *Flammulina velutipes*, *Phyalacriaceae*, and their efficacy for decolorization of methylene blue. *Journal of Environmental Chemical Engineering*, 8(3), 103841.
27. Negahdari B, Darvishi M, Saeedi AA. (2019). Gold nanoparticles and hepatitis B virus. *Artif Cells Nanomed Biotechnol*. 47(1):455-461.
28. Rajeshkumar, S. (2016). Anticancer activity of eco-friendly gold nanoparticles against lung and liver cancer cells. *Journal of Genetic Engineering and Biotechnology*, 14(1), 195–202. <https://doi.org/10.1016/j.jgeb.2016.05.007>
29. Rey, M. W., Ramaiya, P., Nelson, B. A., Brody-Karpin, S. D., Zaretsky, E. J., Tang, M., Lopez de Leon, A., Xiang, H., Gusti, V., Clausen, I. G., Olsen, P. B., Rasmussen, M. D., Andersen, J. T., Jørgensen, P. L., Larsen, T. S., Sorokin, A., Bolotin, A., Lapidus, A., Galleron, N., ... Berka, R. M. (2004). Complete genome sequence of the industrial bacterium *Bacillus licheniformis* and comparisons with closely related *Bacillus* species. *Genome Biology*, 5(10).
30. Rotimi, L., Ojemaye, M. O., Okoh, O. O., Sadimenko, A., & Okoh, A. I. (2019). Synthesis, characterization, antimalarial, antitypanocidal and antimicrobial properties of gold nanoparticle. *Green Chemistry Letters and Reviews*, 12(1), 61–68. <https://doi.org/10.1080/17518253.2019.1569730>
31. Balasubramanian Deepika, Girigoswami Agnishwar and Girigoswami Koyeli . 2023. Antioxidant and anticancer activity of nano lycopene., *Research Journal of Biotechnology*.; Vol. 18(6); 98-104; [doi: https://doi.org/10.25303/1806rjbt98104](https://doi.org/10.25303/1806rjbt98104);
32. Amen Abd , Mahmood N A .Triazole-Anil and Triazol-Azo Reagents (Creation, Spectral Categorization, Scanning Microscopy, Thermal Analysis)., *NeuroQuantology* , 2021; 19(11):84-94 ., DOI Number: 10.14704/nq.2021.19.11.NQ21178
33. S Jawad.; Mahmood N A. Tetrazole Derivatives (Preparation, Organic Analysis, Biotic Evaluation, Nano-Study)., *Egyptian Journal of Chemistry* , 2023, 66 , p:31-40., doi: [10.21608/EJCHEM.2022.152509.6605](https://doi.org/10.21608/EJCHEM.2022.152509.6605)
34. Rulla Sabah, Ahmed saad abbas Fatin F.Al-Kazazz, Salam A.H Al-Ameri , [Investigation on Glucose and levels of Zn and Cu in Sera of Iraqi Males addicted on Methamphetamine or Tramadol](https://doi.org/10.25303/1806rjbt98104) , *Journal of Advanced Sciences and Engineering Technologies: Vol. 3 No. 2 (2020)*
35. Hussein FM. Doped ZnO Nanostructured and their application as photocatalytic: as Review. *Journal of Advanced Sciences and Engineering Technologies*.; 2021 May ;4(1):1-15. Available from: <https://isnra.net/ojs/index.php/jaset/article/view/1>
36. Sabah R. Simultaneous HPLC estimation of Amphetamine and Caffeine abuse drugs in Iraqi human addicts. *Journal of Advanced Sciences and Engineering Technologies* .; 2021 Dec. 13 ;4(1):25-31. Available from: <https://isnra.net/ojs/index.php/jaset/article/view/13>
37. San Diego, K. D. G., Alindayu, J. I. A., & Baculi, R. Q. (2021). Biosynthesis of gold nanoparticles by bacteria from hyperalkaline spring and evaluation of their inhibitory activity against pyocyanin production. *Journal of Microbiology, Biotechnology and Food Sciences*, 2021, 781–787.
38. Sayadi, K., Akbarzadeh, F., Pourmardan, V., Saravani-Aval, M., Sayadi, J., Chauhan, N. P. S., & Sargazi, G. (2021b). Methods of green synthesis of Au NCs with emphasis on their morphology: A mini-review. *Heliyon*, 7(6), e07250.
39. Singh, P. K., & Kundu, S. (2014). Biosynthesis of gold nanoparticles using bacteria. *Proceedings of the National Academy of Sciences India Section B - Biological Sciences*, 84(2), 331–336. <https://doi.org/10.1007/s40011-013-0230-6>
40. Singh, R., Smitha, M. S., & Singh, S. P. (2014). The role of nanotechnology in combating multi-drug resistant bacteria. *Journal of Nanoscience and Nanotechnology*, 14(7), 4745–4756.
41. Soliman, M. K. Y., Salem, S. S., Abu-Elghait, M., & Azab, M. S. (2022). Biosynthesis of Silver and Gold Nanoparticles and Their Efficacy Towards Antibacterial, Antibiofilm, Cytotoxicity, and Antioxidant Activities. *Applied Biochemistry and Biotechnology*,

0123456789. <https://doi.org/10.1007/s12010-022-04199-7>
42. Sonkusre, P. (2020). Specificity of biogenic selenium nanoparticles for prostate cancer therapy with reduced risk of toxicity: an in vitro and in vivo study. *Frontiers in Oncology*, 9, 1541.
 43. Srinath, B. S., Namratha, K., & Byrappa, K. (2018). Eco-friendly synthesis of gold nanoparticles by *Bacillus subtilis* and their environmental applications. *Advanced Science Letters*, 24(8), 5942–5946.
 44. Sriram, M. I., Kalishwaralal, K., & Gurunathan, S. (2012). Biosynthesis of silver and gold nanoparticles using *Bacillus licheniformis*. In *Nanoparticles in biology and medicine* (pp. 33–43). Springer.
 45. Hassan AK, Atiya MA, Luaibi IM. A Green Synthesis of Iron/Copper Nanoparticles as a Catalytic of Fenton-like Reactions for Removal of Orange G Dye. *Baghdad Sci.J* . 2022 Dec. 1 ;19(6):1249. Available from: <https://bsi.uobaghdad.edu.iq/index.php/BSJ/article/view/6508>
 46. Sun, B., Hu, N., Han, L., Pi, Y., Gao, Y., & Chen, K. (2019). Anticancer activity of green synthesised gold nanoparticles from *Marsdenia tenacissima* inhibits A549 cell proliferation through the apoptotic pathway. *Artificial Cells, Nanomedicine, and Biotechnology*, 47(1), 4012–4019.
 47. Sravya M.V.N., Sampath Kumar N.S. Dirisala Vijaya R. , Sai Kiran G.V.S.D., Simhachalam G. 2023. In vitro Assessment of Antibacterial and Antioxidant Activity of *Rhizophora apiculata* leaf extracts ., *Research Journal of Biotechnology*.; Vol. 18(6); 58-65; doi: <https://doi.org/10.25303/1806rjbt058065>;
 48. Balasubramanian Deepika, Girigoswami Agnishwar and Girigoswami Koyeli . 2023. Antioxidant and anticancer activity of nano lycopene., *Research Journal of Biotechnology*.; Vol. 18(6); 98-104; doi: <https://doi.org/10.25303/1806rjbt980104>;
 49. Ayu Y.S. and Kasiamdari R.S . 2023. [Biological Treatment of Naphthol Yellow S and Batik Effluent using *Aspergillus tamaris* and *Aspergillus sclerotiorum*](#)., *Research Journal of Chemistry and Environment*., Vol. 27(6) ,June 2023
 50. Dev Athira S., Hari Neethu and Nair Ananthkrishnan Jayakumaran. Biodegradable natural and synthetic polymers for the development of electrospun nanofibrous scaffolds for various tissue engineering applications., *Research Journal of Biotechnology*., 2023, Vol. 18(6); 115-131; doi: <https://doi.org/10.25303/1806rjbt1150131>;
 51. Jessica María Soria Villanes, Mónica Evencia Poma Vivas, Camila Alejandra Traverso Castillo, Michelle Norma Antonio . (2023). El microbio, camino biológico a través de la especie humana., *Boletín de Malariología y Salud Ambiental*., Vol. 63, Núm. 2 (2023), p: 330-337.
 52. Ruth Katherine Mendivel Geronimo, Jessica Coronel Capani, Oscar Lucio Ninamango Solis, Juan de Dios Aguilar Sánchez, Ana María Enriquez Chauca .(2023). Rasgos antropogénicos de los ciclos zoonóticos en el Perú ., *Boletín de Malariología y Salud Ambiental*.,Vol. 63, Núm. 2 (2023), p:338-349
 53. Osmer Campos-Ugaz, Patricia Campos Olazábal, Ronald M. Hernández, Silvia Georgina Aguinaga Doig, Janeth Benedicta Falla Ortiz, Emma Margarita Wong Fajardo, Lydia Mercedes Morante Becerra. (2023). Investigación formativa en epidemiología crítica de la enfermedades infecciosas en Latinoamérica 2010 al 2020 ., *Boletín de Malariología y Salud Ambiental*., Vol. 63, Núm. 2 (2023), p: 350-360.
 54. Xie, P., Qi, Y., Wang, R., Wu, J., & Li, X. (2019). Aqueous gold nanoparticles generated by AC and pulse-power-driven plasma jet. *Nanomaterials*, 9(10), 1488.
 55. Yuan, X., Ge, L., Zhou, H., & Tang, J. (2023). Size, composition, and surface capping-dependent catalytic activity of spherical gold nanoparticles. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 287, 122082.
 56. Zhao, H., Shi, K., Zhang, C., Ren, J., Cui, M., Li, N., Ji, X., & Wang, R. (2022). Spherical COFs decorated with gold nanoparticles and multiwalled carbon nanotubes as signal amplifier for sensitive electrochemical detection of doxorubicin. *Microchemical Journal*, 182, 107865.